

Article ID: 9204
DOI: 10.5586/asbp.9204

Publication History
Received: 2022-04-11
Accepted: 2022-12-13
Published: 2023-03-17

Handling Editor
Przemysław Wojtaszek; Adam Mickiewicz University, Poznań, Poland; <https://orcid.org/0000-0002-4484-1536>

Authors' Contributions
CZS and LYY designed the research; MYL, XFS and JSZ performed the experiments; MYL, CZS and JSZ wrote the paper; HNC, XLL, and XYZ provided experimental assistance; all authors revised the manuscript

Funding
This work was supported by the National Natural Science Foundation of China (31801877) and the Hebei Province High Education Science and Technology Research Project (BJ2018002).

Competing Interests
No competing interests have been declared.

Copyright Notice
© The Author(s) 2023. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits redistribution, commercial and noncommercial, provided that the article is properly cited.

RESEARCH PAPER

CsCLE3 delays female flower anthesis in cucumber

Mingyue Lei, Xiaofei Song, Jinshuang Zheng, Chengzhen Sun ^{*}, Liying Yan

Hebei Key Laboratory of Horticultural Germplasm Excavation and Innovative Utilization, College of Horticulture Science and Technology, Hebei Normal University of Science and Technology, Qinhuangdao 066004, China

^{*} To whom correspondence should be addressed. Email: chengzhensun@126.com

Abstract

A corolla opening is a necessary process affecting the quality of fruits and market competitiveness in cucumber (*Cucumis sativus*). In the previous paper, we identified a unique cucumber line ('6457') that possesses extra-long ovaries and shows a delayed corolla opening when nutrient supplies are abundant. We also previously showed that the expression of *CsCLE3* (*Csa4G627800*) is correlated with the delayed opening of the female corolla. Here, we investigated the function of *CsCLE3* in cucumber by conducting transgenic experiments and phenotypic analysis. The results showed that the expression of *CsCLE3* in the extra-long ovary was significantly lower than in the typical ovary. In *CsCLE3*-overexpressed plants, the capacity to produce extra-long ovaries was lost, and the average rates of the extra-long ovary and the extra-long ovary plant were both 0%. In the *CsCLE3* knockout plants obtained by the CRISPR/Cas9 system, the average extra-long ovary and extra-long ovary plant rates were significantly higher- 66.67% and 100%, respectively. Our study proved a negative regulating corolla opening time factor and provides new insight into the molecular basis of cucumber reproduction, producing fruits with flowers remaining on the tip.

Keywords

CsCLE3; corolla opening; *Cucumis sativus*; gene function; extra-long ovary

1. Introduction

An opening of the corolla is an essential stage in higher plants' life cycle, regulated by a complex set of hormonal and external environmental factors (Larsson et al., 1998; Prudencio et al., 2020). Several pathways have been identified to regulate flowering in the model plant *Arabidopsis thaliana* (He, 2012), including photoperiod, vernalization, gibberellin, ambient temperature, and age-related pathways.

Polypeptide hormones are important signal transduction molecules in plants and play a crucial role in growth and development (Song et al., 2013). CLE family is a relatively large class of polypeptide signaling molecules in plants (Fletcher et al., 1999; Opsahl-Ferstad et al., 1997; Strabala et al., 2006). The *A. thaliana* CLE family contains 32 members with conserved CLE motifs and signal peptides (Chu et al., 2006). *In vitro*, when the small peptide (synthesized from the CLE motif of *CLV3*) applies, *A. thaliana* presents a phenotype of gene overexpression, indicating that *CLV3* is a secretory peptide (Fiers et al., 2005). *CLV3* is mainly expressed in the central region of *A. thaliana* shoot apical meristem (SAM) as a dynamic apoplast signal and regulates the stem cell development through the *CLV3*-*WUS* feedback loop pathway (Brand et al., 2000; Schoof et al., 2000; Shinohara & Matsubayashi, 2013). The root of functional deletion mutant *cle40*-En becomes shorter and tilted to the left (Hobe et al., 2003). CLE40 can bind to the receptor ACR4, inhibit the expression of *WOX5*, and maintain the balance between proliferation and differentiation of apical root meristem (RAM) stem cells through *CLE40*-*WOX5* feedback regulatory loop (Stahl et al., 2009).

It is also found that *CLE41* and *CLE44* could further enhance the proliferation of stem cells by promoting the expression of *WOX4* (Qiang et al., 2013). *CLE8-WOX8* signal transduction pathway is involved in the early development of embryo and endosperm in *A. thaliana* (Fiume & Fletcher, 2012). *AtCLE25* positively regulates water transport from roots to shoots (Takahashi et al., 2018). *CLE3* acts as a root-derived long-distance signal that regulates shoots' systemic acquired resistance (SAR) by interaction with *WRKY33* (Ma et al., 2020, 2022).

The CLE family is a relatively large class of polypeptide signaling molecules in plants, and overexpressing *CLE19*, *CLE21*, or *CLE25* delays the development of rosettes (Strabala et al., 2006). In *A. thaliana*, *AtCLE16*, *AtCLE17*, and *AtCLE22* display high promoter activity in sepals and petals at different flower development stages (Jun et al., 2010). However, there is little research on the functions of plant peptides *CLE3* in the development of flowers.

A corolla opening is an important process affecting the quality of fruits and market competitiveness in cucumber. We introduced a unique cucumber line, '6547', that possesses extra-long ovaries and shows delayed corolla opening when nutrient supplies are abundant. The extra-long ovary reaches the commercial harvesting stage 2–3 d after flowering while the corolla remains fresh. The obtained marketable cucumber fruits from the two types of the ovary are indistinguishable in length. However, the fruits from extra-long ovaries are straight and have flowers remaining on the tip, which is regarded as more attractive and preferred in the fresh market (Sun et al., 2016). Interestingly, by transcriptome analysis, we have also found that the expression of *CsCLE3* (*Csa4G627800*) negatively correlates with the formation of an extra-long ovary (Sun et al., 2016), suggesting that the gene regulates extra-long ovary formation.

Here, we investigate the function of *CsCLE3* and reveal the mechanism of the delayed female corolla opening in cucumbers by conducting transgenic experiments and phenotypic analysis.

2. Material and methods

2.1. Plant materials and treatments

The cucumber (*Cucumis sativus*) line '6457' was grown in the greenhouse of Hebei Normal University of Science and Technology under two cultivation treatments described in the previous study (Sun et al., 2016).

2.2. The extra-long ovary rate and extra-long ovary plant rate analysis

The extra-long ovary rate and extra-long ovary plant rate were calculated using the following formulas:

$$\text{Extra-long ovary rate (\%)} = \frac{\text{number of extra-long ovaries}}{\text{total number of ovaries}} \times 100\%$$

$$\text{Extra-long ovary plant rate (\%)} = \frac{\text{number of plants that produced extra-long ovaries}}{\text{total number of plants in the same stadium}} \times 100\%$$

2.3. Quantitative real-time PCR (qRT-PCR)

Based on color and shape, corolla development can be divided into four stages: green bud, green-yellow bud, yellow bud, and flowering. In the typical ovary, 1 DAL (T1), 3 DAL (T3), 4 DAL (T4), and 5 DAL (T5) indicated the green bud, green-yellow bud, yellow bud, and flowering stages, respectively. In contrast, in the extra-long ovary, these four stages typically corresponded to 1 DAL (EL1), 3 to 5 DAL (EL3, EL4, and EL5), 7 to 8 DAL (EL7 and EL8), and 9 DAL (EL9), respectively. Female corollas at different developmental stages under typical ovary and extra-long ovary conditions were frozen in liquid nitrogen and stored at -80°C for total RNA extraction (Waryoung, China, <http://www.huayueyang.com>). First-strand cDNA was synthesized and used as a template for qPCR with an SYBR Premix Taq Mix (Takara) on Applied Biosystems

7500 real-time PCR system. The thermal conditions for real-time PCR were 95 °C for 10 min (denaturation) followed by 40 cycles at 95 °C for 15 s and 60 °C for 1 min.

Alpha-tubulin (*TUA*, AJ715498) and ubiquitin-like protein (*UBI*, AF104391) were used as candidates for reference genes (Wan et al., 2009). The expression profiles and stability of cucumber ovary at different physiological developmental stages for two reference genes were analyzed according to the literature (Liu et al., 2020), using three different methods with BestKeeper (Pfaffl et al., 2004), geNorm (Vandesompele et al., 2002), and NormFinder (Andersen et al., 2004) software packages. Primer efficiencies and standard deviations were calculated using qBase software (version 1.3.5) (Hellemans et al., 2007) on a standard curve generated using a five-fold dilution series of one sample over at least six dilution points measured in triplicate. Three biological and three technical repeats were conducted for each gene. Statistical analysis was conducted with two-tailed Student's t-tests (* $P < 0.05$, ** $P < 0.01$). Primers are listed in Table S1.

2.4. Cucumber transformation

The full-length coding sequence (CDS) of *CsCLE3* was inserted into the pCAMBIA1305.1 (Novagen, USA) vector to generate the Pro35S: *CsCLE3* overexpression vector. Two target sequences (19 bp) located in the exon were used to construct the CRISPR/Cas9 knockout vector (pKSE402 with GFP fluorescent screening marker, pCBC-DT1T2 as an intermediate vector). The recombinant knockout vector was transformed into *Agrobacterium tumefaciens* EHA105. The detailed cucumber transformation protocol can be found in the previous study (Hu et al., 2017). Cotyledons were infected by EHA105 (in infection fluid with OD600 = 0.2–0.3) under negative pressure. After three days of co-culture in darkness, explants were transferred to a bud differentiation medium with timentin (200 µg/L) under light/8 h dark at 26 °C for 3–4 weeks (Ding et al., 2015). Then the buds with the GFP marker were selected, excised from an explant, and transferred to a rooting medium (Ding et al., 2015). The homozygous T1 mutants without vector (GFP-free) were identified from T0 transgenic lines for further phenotype observation and data statistics. Primers are listed in Table S1.

3. Results

3.1. Expression patterns of *CsCLE3*

To check the stability of the candidate reference genes *TUA* and *UBI* in a cucumber ovary at physiological development stages, we used RT-qPCR. We observed the expression of the two reference genes at the T1, T3, and EL7. The results showed that the Ct values of the two candidates were between 15 and 21 at T1, T3, and EL7 (Figure S1A). The geNorm, NormFinder, and BestKeeper stability analysis showed that *TUA* and *UBI* are stable reference genes (Figure S1B-E).

In this study, the relative expression levels of *CsCLE3* in the female flowers at different developmental stages under the typical ovary and the extra-long ovary conditions of '6457' were measured by qRT-PCR. We found that *CsCLE3* had similar expression patterns both under the typical ovary and the extra-long ovary conditions based on the data of transcriptome analysis and qRT-PCR (Figure 1). The expression of *CsCLE3* increased significantly at the stage of yellow bud (N7/S7/S8) and then decreased significantly. However, the expression level of *CsCLE3* in the extra-long ovary was significantly lower than that of the typical ovary during the female corolla opening.

3.2. *CsCLE3* Negatively regulates extra-long ovary formation in cucumber

To identify the biological functions of *CsCLE3* in cucumber, the Pro35S: *CsCLE3* overexpression vector contained the full-length CDS of *CsCLE3* driven by the CaMV 35S promoter, was transformed into '6457'. The representative line *CsCLE3-oe-11* was used for further characterization. Our data showed that the overexpression of *CsCLE3* inhibited the formation of extra-long ovaries. Both the average extra-long ovary rate and the extra-long ovary plant rate were 0% (Figure 2B, Table 1). The overexpressed

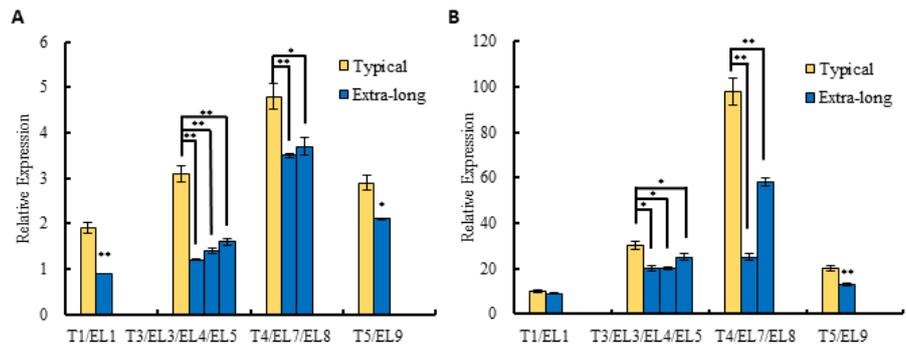


Figure 1 Relative expression of *CsCLE3* at different stages of corolla opening in ‘6457’.

(A) Transcriptome analysis (Sun et al., 2016). (B) The qRT-PCR analysis. Two-tailed Student’s t-tests were used (* $P < 0.05$, ** $P < 0.01$). The four stages of corolla development in the typical ovary (T) and extra-long ovary (EL) at which samples were collected for analysis: green bud, green-yellow bud, yellow bud, and flowering. In the typical ovary, 1 DAL (T1), 3 DAL (T3), 4 DAL (T4), and 5 DAL (T5) indicated the green bud, green-yellow bud, yellow bud, and flowering stages, respectively. In contrast, in the extra-long ovary, these four stages typically corresponded to 1 DAL (EL1), 3 to 5 DAL (EL3, EL4, and EL5), 7 to 8 DAL (EL7 and EL8), and 9 DAL (EL9), respectively.

Table 1 Extra-long ovary rate and extra-long ovary plant rate for different lines.

Line	Extra-long ovary rate (%)	Extra-long ovary plant rate (%)
6457	29.38	24.05
<i>Cscl3-oeor-11</i>	0	0
<i>Cscl3-cr-2</i>	66.67	100.00

line *Cscl3-oe-11* mostly entered anthesis at 5 d after labeling (DAL; labeling was done when an ovary became visible) (Figure 2C).

To confirm the function of *CsCLE3* in cucumber, we constructed *CsCLE3*-knockout lines in ‘6457’ by the CRISPR-Cas9 gene-editing system. The two targets are located in the exon. One loss-of-function mutant line (*Cscl3-cr-2*) was selected for further characterization (Figure 2A). The knockout of *CsCLE3* in ‘6457’ could have enhanced the extra-long ovary phenotype. The average extra-long ovary rate was 66.67%, and the average extra-long ovary plant rate was 100% (Figure 2B, Table 1). According to the quantification of the days to blooming, we found that the extra-long ovary of *Cscl3-cr-2* mostly entered anthesis at 8–10 DAL, which was 4-d to 5-d delayed in comparison to the typical ovary (Figure 2C).

For the typical (*Cscl3-oe-11*) and the extra-long ovary (*Cscl3-cr-2*) of ‘6457’, the average ovary length on the day of the corolla opening was 5.4 cm and 11.8 cm, respectively (Figure 2D). However, among the three lines, the ovary growth rates and length at the marketable stage of the two types of ovaries correspond to each other (Figure 2E).

4. Discussion

A corolla opening is an evolutionary breakthrough in the reproduction of higher plants (van Doorn & Kamdee, 2014). Nevertheless, the molecular mechanism of a corolla opening remains largely unknown, mainly due to the lack of mutants or lines that display appreciable changes during a corolla opening. In cucumber, the time of the female flower anthesis and corolla durability is a vital appearance quality trait that requires extensive investigation. For most cucumber cultivars, this process takes 4d to 5d and is insensitive to nutrient conditions (Sun et al., 2016). In cucumber inbred line ‘6547’, the corolla opening of flowers with the extra-long ovary was 4–5 days delayed compared to flowers with the typical ovary. However, the female flower anthesis’s

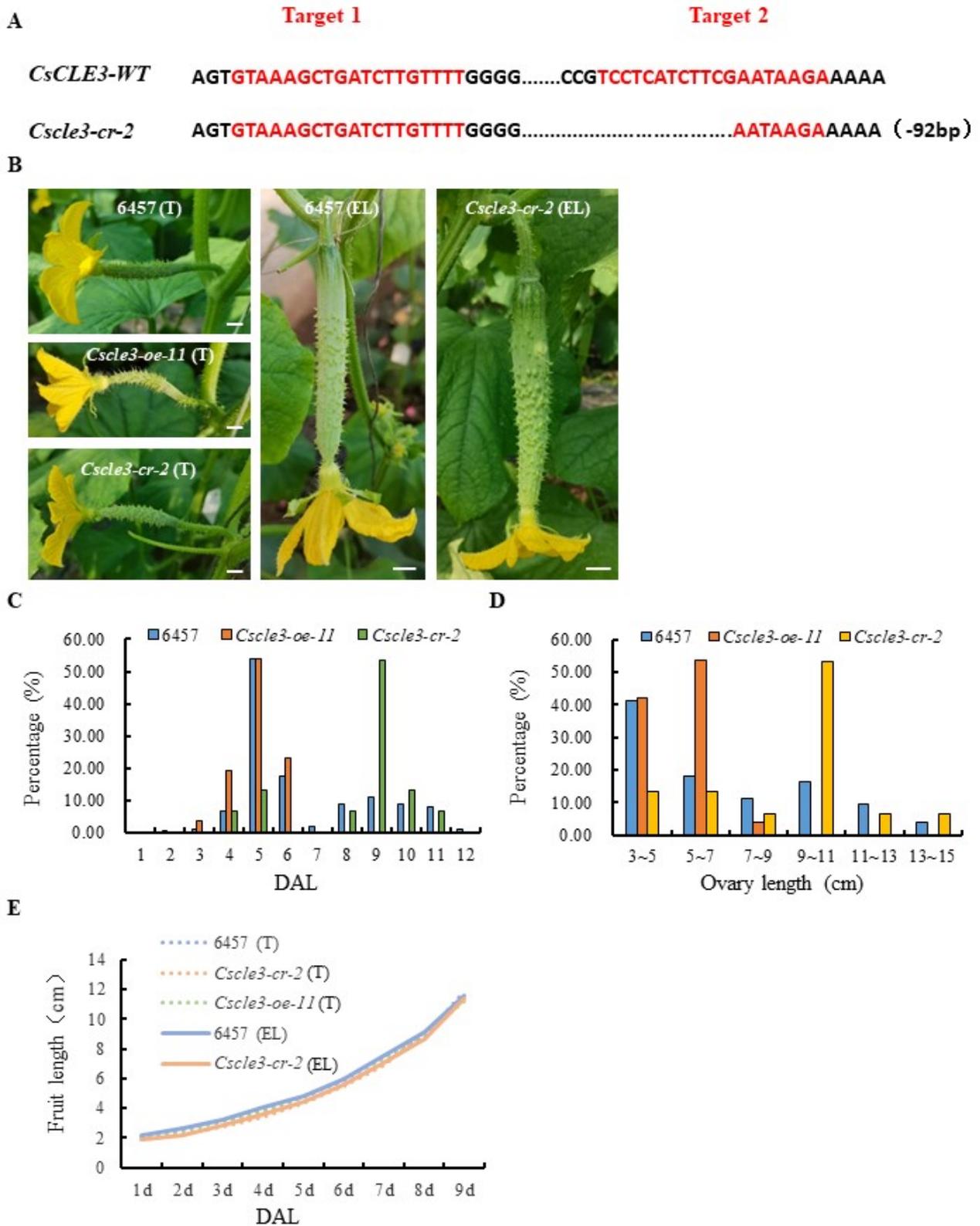


Figure 2 Morphological characterization of the transformation lines.

(A) Mutation form of T1 transgenic *Cscl3-cr-2* line by CRISPR/Cas9 system. (B) Ovary and corolla phenotype at the day of corolla open in ‘6457’, *Cscl3-oe-11*, and *Cscl3-cr-2*. Quantification of the days to anthesis (C), ovary length at the day of corolla open (D), and development rates of ovary length in ‘6457’, *Cscl3-oe-11*, and *Cscl3-cr-2* (E). Scale bar = 1 cm.

main genes and molecular mechanism remain unknown. In this study, we found through transgenic experiments that *CsCLE3* delays the female flower anthesis in cucumbers. These findings deepen our knowledge of the female flower anthesis and provide new insight into the basis of molecular cucumber reproduction, producing fruits with flowers remaining on the tip.

5. Supplementary material

The following supplementary material is available for this article:

Table S1. The sequence of primers.

Figure S1. Expression of two candidate reference genes and stability of cucumber ovary in different physiological developmental stages.

References

- Andersen, C. L., Jensen, J. L., & Orntoft, T. F. (2004). Normalization of real-time quantitative reverse transcription-PCR data: A model-based variance estimation approach to identify genes suited for normalization, applied to bladder and colon cancer data sets. *Cancer Research*, *64*(15), 5245–5250. <https://doi.org/10.1158/0008-5472.CAN-04-0496>
- Brand, U., Fletcher, J. C., Hobe, M., Meyerowitz, E. M., & Simon, R. (2000). Dependence of stem cell fate in *Arabidopsis* on a feedback loop regulated by CLV3 activity. *Science*, *289*(5479), 617–619. <https://doi.org/10.1126/science.289.5479.617>
- Chu, H. W., Qian, Q., Liang, W. Q., Yin, C. S., Tan, H. X., Yao, X., Yuan, Z., Yang, J., Huang, H., Luo, D., Ma, H., & Zhang, D. B. (2006). The *FLORAL ORGAN NUMBER4* gene encoding a putative ortholog of *Arabidopsis* CLAVATA3 regulates apical meristem size in rice. *Plant Physiology*, *142*(3), 1039–1052. <https://doi.org/10.1104/pp.106.086736>
- Ding, L., Yan, S. S., Jiang, L., Zhao, W. S., Ning, K., Zhao, J. Y., Liu, X. F., Zhang, J., Wang, Q., & Zhang, X. L. (2015). *HANABA TARANU* (*HAN*) bridges meristem and organ primordia boundaries through *PINHEAD*, *JAGGED*, *BLADE-ON-PETIOLE2*, and *CYTOKININ OXI-DASE 3* during flower development in *Arabidopsis*. *PLoS Genetics*, *11*(9), Article e1005479. <https://doi.org/10.1371/journal.pgen.1005479>
- Fiers, M., Golemic, E., Xu, J., van der Geest, L., Heidstra, R., Stiekema, W., & Liu, C. M. (2005). The 14-amino acid CLV3, CLE19, and CLE40 peptides trigger consumption of the root meristem in *Arabidopsis* through a *CLAVATA2*-dependent pathway. *Plant Cell*, *17*(9), 2542–2553. <https://doi.org/10.1105/tpc.105.034009>
- Fiume, E., & Fletcher, J. C. (2012). Regulation of *Arabidopsis* embryo and endosperm development by the polypeptide signaling molecule CLE8. *Plant Cell*, *24*(3), 1000–1012. <https://doi.org/10.1105/tpc.111.094839>
- Fletcher, J. C., Brand, U., Running, M. P., Simon, R., & Meyerowitz, E. M. (1999). Signaling of cell fate decisions by CLAVATA3 in *Arabidopsis* shoot meristems. *Science*, *283*(5409), 1911–1914. <https://doi.org/10.1126/science.283.5409.1911>
- He, Y. H. (2012). Chromatin regulation of flowering. *Trends in Plant Science*, *17*(9), 556–562. <https://doi.org/10.1016/j.tplants.2012.05.001>
- Hellems, J., Mortier, G., De Paepe, A., Speleman, F., & Vandesompele, J. (2007). qBase relative quantification framework and software for management and automated analysis of real-time quantitative PCR data. *Genome Biology*, *8*, R19. <https://doi.org/10.1186/gb-2007-8-2-r19>
- Hobe, M., Müller, R., Grünwald, M., Brand, U., & Simon, R. (2003). Loss of CLE40, a protein functionally equivalent to the stem cell restricting signal CLV3, enhances root waving in *Arabidopsis*. *Development Genes and Evolution*, *213*(8), 371–381. <https://doi.org/10.1007/s00427-003-0329-5>
- Hu, B., Li, D., Liu, X., Qi, J., Gao, D., Zhao, S., Huang, S., Sun, J., & Yang, L. (2017). Engineering non-transgenic gynocercous cucumber using an improved transformation protocol and optimized CRISPR/Cas9 system. *Molecular Plant*, *10*(12), 1575–1578. <https://doi.org/10.1016/j.molp.2017.09.005>
- Jun, J., Fiume, E., Roeder, A. H., Meng, L., Sharma, V. K., Osmont, K. S., Baker, C., Ha, C. M., Meyerowitz, E. M., Feldman, L. J., & Fletcher, J. C. (2010). Comprehensive analysis of CLE polypeptide signaling gene expression and overexpression activity in *Arabidopsis*. *Plant Physiology*, *154*(4), 1721–1736. <https://doi.org/10.1104/pp.110.163683>
- Larsson, A. S., Landberg, K., & Meeks-Wagner, D. R. (1998). The *TERMINAL FLOWER2* (*TFL2*) gene controls the reproductive transition and meristem identity in *Arabidopsis thaliana*. *Genetics*, *149*(2), 597–605. <https://doi.org/10.1093/genetics/149.2.597>
- Liu, L., Han, H., Li, Q. X., Chen, M., Zhou, S. Q., Wang, H., & Chen, L. B. (2020). Selection and validation of the optimal panel of reference genes for RT-qPCR analysis in the

- developing rat cartilage. *Frontiers in Genetics*, 11, 1664–8021.
<https://doi.org/10.3389/fgene.2020.590124>
- Ma, D., Endo, S., Betsuyaku, E., Fujiwara, T., Betsuyaku, S., & Fukuda, H. (2022). Root-specific *CLE3* expression is required for *WRKY33* activation in *Arabidopsis* shoots. *Plant Molecular Biology*, 108(3), 225–239. <https://doi.org/10.1007/s11103-021-01234-9>
- Ma, D., Endo, S., Betsuyaku, S., Shimotohno, A., & Fukuda, H. (2020). *CLE2* regulates light-dependent carbohydrate metabolism in *Arabidopsis* shoots. *Plant Molecular Biology*, 104, 561–574. <https://doi.org/10.1007/s11103-020-01059-y>
- Opsahl-Ferstad, H. G., Le, D. E., Dumas, C., & Rogowsky, P. M. (1997). *ZmEsr*, a novel endosperm-specific gene expressed in a restricted region around the maize embryo. *Plant Journal*, 12(1), 235–246. <https://doi.org/10.1046/j.1365-3113X.1997.12010235.x>
- Pfaffl, M. W., Tichopad, A., Prgomet, C., & Neuvians, T. P. (2004). Determination of stable housekeeping genes, differentially regulated target genes, and sample integrity: BestKeeper–Excel-based tool using pair-wise correlations. *Biotechnology Letters*, 26, 509–515. <https://doi.org/10.1023/b:bile.0000019559.84305.47>
- Prudencio, A. S., Hoerberichts, F. A., Dicenta, F., Martínez-Gómez, P., & Sánchez-Pérez, R. (2020). Identification of early and late flowering time candidate genes in endodormant and ecodormant almond flower buds. *Tree Physiology*, 41(4), 589–605.
<https://doi.org/10.1093/treephys/tpaa151>
- Qiang, Y., Wu, J., Han, H., & Wang, G. (2013). CLE peptides in vascular development. *Journal of Plant Ecology*, 55(4), 389–394. <https://doi.org/10.1111/jipb.12044>
- Schoof, H., Lenhard, M., Haecker, A., Mayer, K. F., Jürgens, G., & Laux, T. (2000). The stem cell population of *Arabidopsis* shoot meristems is maintained by a regulatory loop between the *CLAVATA* and *WUSCHEL* genes. *Cell*, 100(6), 635–644.
[https://doi.org/10.1016/S0092-8674\(00\)80700-X](https://doi.org/10.1016/S0092-8674(00)80700-X)
- Shinohara, H., & Matsubayashi, Y. (2013). Chemical synthesis of *Arabidopsis* CLV3 glycopeptide reveals the impact of hydroxyproline arabinosylation on peptide conformation and activity. *Plant Cell Physiology*, 54(3), 369–374.
<https://doi.org/10.1093/pcp/pcs174>
- Song, X. F., Guo, P., Ren, S. C., Xu, T. T., & Liu, C. M. (2013). Antagonistic peptide technology for functional dissection of *CLV3/ESR* genes in *Arabidopsis*. *Plant Physiology*, 161(3), 1076–1085. <https://doi.org/10.1104/pp.112.211029>
- Stahl, Y., Wink, R. H., Ingram, G. C., & Simon, R. (2009). A signaling module controlling the stem cell niche in *Arabidopsis* root meristems. *Current Biology*, 19(11), 909–914.
<https://doi.org/10.1016/j.cub.2009.03.060>
- Strabala, T. J., O'Donnell, P. J., Smit, A. M., Ampomah-Dwamena, C., Martin, E. J., Netzler, N., Nieuwenhuizen, N. J., Quinn, B. D., Foote, H. C. C., & Hudson, K. R. (2006). Gain-of-function phenotypes of many *CLAVATA3/ESR* genes, including four new family members, correlate with tandem variations in the conserved *CLAVATA3/ESR* domain. *Plant Physiology*, 140(4), 1331–1344. <https://doi.org/10.1104/pp.105.075515>
- Sun, C. Z., Li, Y. Q., Zhao, W. S., Song, X. F., Lu, M., Li, X. L., Li, X. X., Liu, R. Y., Yan, L. Y., & Zhang, X. L. (2016). Integration of hormonal and nutritional cues orchestrates progressive corolla opening. *Plant Physiology*, 171(2), 1209–1229.
<https://doi.org/10.1104/pp.16.00209>
- Takahashi, F., Suzuki, T., Osakabe, Y., Betsuyaku, S., Kondo, Y., Dohmae, N., Fukuda, H., Yamaguchi-Shinozaki, K., & Shinozaki, K. (2018). A small peptide modulates stomatal control via abscisic acid in long-distance signaling. *Nature*, 556, 235–238.
<https://doi.org/10.1038/s41586-018-0009-2>
- Vandesompele, J., Preter, K. D., Pattyn, F., Poppe, B., Roy, N. V., Paepe, A. D., & Speleman, F. (2002). Accurate normalization of real-time quantitative RT-PCR data by geometric averaging of multiple internal control genes. *Genome Biology*, 3(7), RESEARCH0034.
<https://doi.org/10.1186/gb-2002-3-7-research0034>
- van Doorn, W. G., & Kamdee, C. (2014). Flower opening and closure: an update. *Journal of Experimental Botany*, 65, 5749–5757. <https://doi.org/10.1093/jxb/eru327>
- Wan, H. J., Zhao, Z. G., Qian, C. T., Sui, Y. H., Malik, A. A., & Chen, J. F. (2009). Selection of appropriate reference genes for gene expression studies by quantitative real-time polymerase chain reaction in cucumber. *Analytical Biochemistry*, 399(2), 257–261.
<https://doi.org/10.1016/j.ab.2009.12.008>